

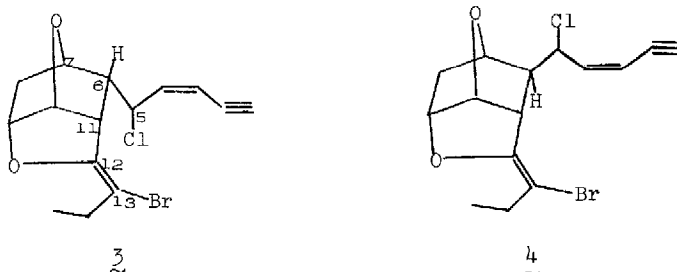
C<sub>15</sub>-HALOGENATED COMPOUNDS FROM THE HAWAIIAN MARINE ALGA  
LAURENCIA NIDIFICA. VI. THE ISOMANEONENES

H. H. Sun, S. M. Waraszkiewicz, and K. L. Erickson\*  
 Jeppson Laboratory, Clark University  
 Worcester, Massachusetts 01610

(Received in USA 17 August 1976; received in UK for publication 4 October 1976)

We recently reported the structures of the maneonenes, three nonterpeneoid C<sub>15</sub>-halogenated ethers from the green variety of Laurencia nidifica.<sup>1</sup> Two other components of this extract, isomaneonene-A (1) and isomaneonene-B (2), have now been characterized in addition to a fourth maneonene, cis-maneonene-C (3).

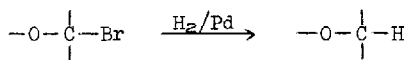
cis-Maneonene-C (3) is the C<sub>6</sub>-endo isomer of cis-maneonene-A (4).<sup>1</sup> This structural assignment was made by application of the same spectral and chemical methods described for the other maneonenes.<sup>1,2</sup> The difference in stereochemistry at C<sub>6</sub> is clearly evident from the pmr where J<sub>6,7</sub> = 4.5 and J<sub>6,11</sub> = 9.5 Hz in 3 while J<sub>6,7</sub> = 0 and J<sub>6,11</sub> = 1.5 Hz in 4.<sup>3</sup>



Isomaneonene-A (1) and isomaneonene-B (2) both analyzed for C<sub>15</sub>H<sub>16</sub>Br<sub>2</sub>O<sub>2</sub> by high resolution and field desorption mass spectroscopy, and their spectral data indicated a close relationship with cis-maneonene-C (3). The cis-enyne function was established by uv ( $\lambda_{\text{max}}^{\text{EtOH}}$  229 nm,  $\epsilon = 14,500$  for 1 and 228 nm,  $\epsilon = 13,500$  for 2), ir (CCl<sub>4</sub>, 3300 and 2970 cm<sup>-1</sup>), pmr and cmr (Tables I and II, C<sub>1</sub>-C<sub>4</sub>). The infrared showed no hydroxyl or carbonyl absorption, but strong bands in the 1000-1100 cm<sup>-1</sup> region indicated that the oxygen moieties were present as ethers. Unlike the maneonenes, neither 1 nor 2 showed vinyl ether absorption. That the cis-enyne group is the only multiply-bonded system in the molecule is established by pmr and cmr (Tables I and II), and therefore isomaneonene-A (1) and isomaneonene-B (2) must be tetracyclic.

Decoupling experiments in the pmr established a proton sequence for C<sub>1</sub>-C<sub>11</sub> and specific hydrogen decoupling in the cmr verified that C<sub>7</sub>, C<sub>9</sub>, and C<sub>10</sub> were oxygen-bearing carbons (Figure I). The four remaining carbons are two deshielded quaternary centers with an ethyl group attached (Tables I and II, C<sub>12</sub>-C<sub>15</sub>) (Figure I). The chemical shift values of the quaternary carbons C<sub>12</sub> and C<sub>13</sub> indicate that the two bromine atoms and the fourth oxygen bond are attached to these carbons. Furthermore, one of these quaternary centers bears both a bromine and an oxygen for when both bromines are reductively removed, the fully saturated diether obtained (5, Scheme I)<sup>2</sup> displays four, rather than

three, oxygen methines in the pmr indicative of the following transformation:



This same ether bond is cleaved when 1 or 2 is treated with Na/NH<sub>3</sub>. Hydrogenation of the resulting olefin gives rise to a secondary alcohol (6, Scheme I). The other ether bonds are unaffected.

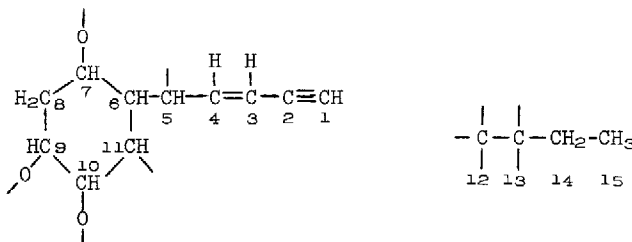


Figure I<sup>4</sup>

With both an oxygen and a bromine on one quaternary center, the second bromine must then be attached to the adjacent quaternary carbon. The C<sub>12</sub>-C<sub>15</sub> moiety shown in Figure I may then be represented by either of two structures:



That a is the correct structure is shown by the mass spectra of secondary alcohol 6 and its corresponding ketone 7 where both bromines and the ether link to the bromine-bearing carbon have been replaced by hydrogens. These materials lose butyl and ethyl fragments on electron impact. Had b been the correct structure, reduction would have given rise to a propyl, not an ethyl, side chain.

Attaching a to the main chain in Figure I at C<sub>5</sub> and C<sub>11</sub> and closing the ether functions so as to exclude an epoxide link (for which there is no evidence), results in two possible skeletal structures for isomaneonene-A and isomaneonene-B. An examination of the molecular models of these two structures shows that only one skeleton will fit the data, that represented by 1 and 2 (Scheme I). It can be seen that these structures, differing only in stereochemistry at C<sub>13</sub>, are identical to the maneonene system with an extra carbocyclic ring formed between C<sub>5</sub> and C<sub>13</sub> and the second halogen attached at C<sub>12</sub> rather than C<sub>5</sub>.

The distinction between isomaneonene-A (1) and isomaneonene-B (2) is readily made on the basis of their pmr spectra. In 1 the bromine atom on C<sub>13</sub> is cis to H<sub>5</sub> and trans to the enyne function and H<sub>4</sub>, while the opposite situation obtains in 2. The deshielding effect of this bromine is shown where H<sub>5</sub> occurs 0.80 ppm further downfield in 1 than in 2 while H<sub>4</sub> is 0.68 ppm further downfield in 2 than in 1. H<sub>6</sub> and H<sub>11</sub> also occur substantially further downfield when the C<sub>13</sub>-bromine is cis to them (Tables I and II).

The cis relationship of H<sub>5</sub> and H<sub>6</sub> in 1 and 2 is established by their large coupling constants (10 and 9 Hz respectively), comparable to J<sub>6,11</sub> (10 Hz). From the models the dihedral angles of H<sub>5</sub>,H<sub>6</sub> and H<sub>6</sub>,<sub>11</sub> are identical. The stereochemistry at all other centers is fixed by virtue of the fused ring systems.

It should be noted that the green variety of L. nidifica from which this varied group of secondary metabolites has been isolated<sup>1,5</sup> may itself consist of several different varieties.

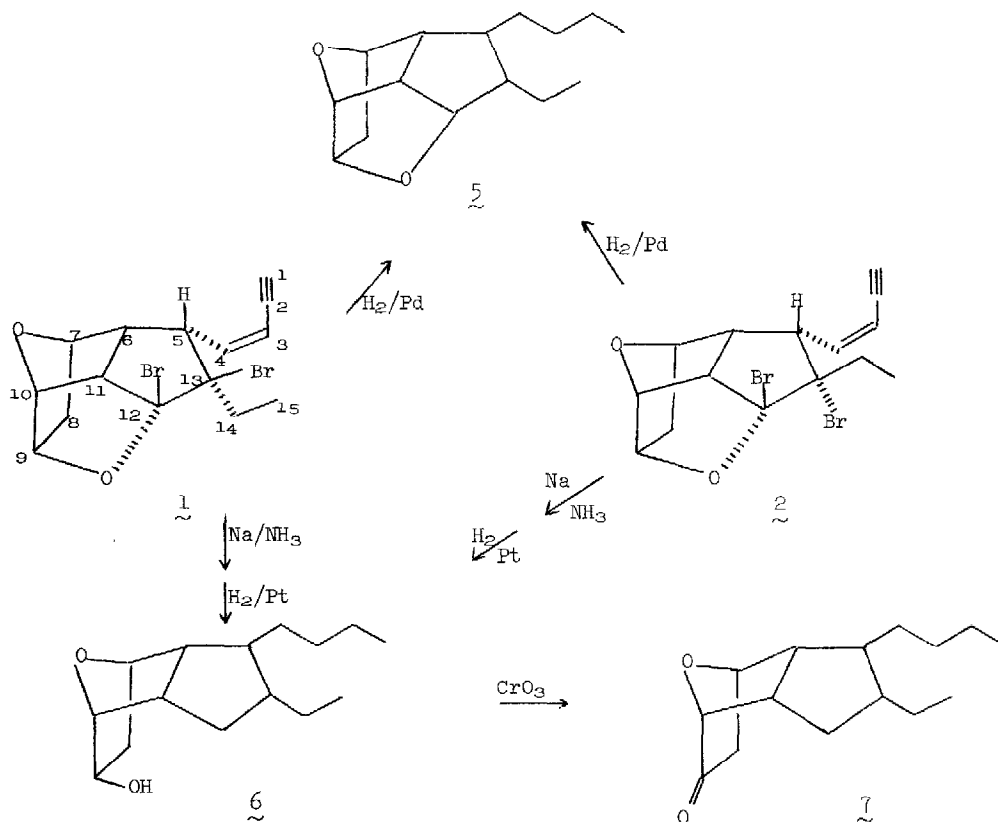
Table I.  $^{13}\text{C}^{\text{a}}$  and  $^1\text{H}^{\text{b}}$  NMR Data for Isomaneonene-A (1)

Carbon No.	$^{13}\text{C}$ $\delta$	$^1\text{H}$ $\delta$	Multiplicity, J (Hz)
1	84.7	2.72	d, $J_{1,3} = 2$
2	79.5 <sup>c,d</sup>	-	-
3	109.2	4.98	dd, $J_{1,3} = 2, J_{3,4} = 10.5$
4	144.5	5.66	dd, $J_{3,4} = 10.5, J_{4,5} = 8$
5	49.7	4.33	dd, $J_{4,5} = 8, J_{5,6} = 10$
6	50.5	3.38	sextet, $J_{5,6} = J_{6,11} = 10, J_{6,7} = 5$
7	77.6	4.19	t, $J_{6,7} = J_{7,8_{\text{exo}}} = 5$
8	41.4	$\begin{cases} 1.15 \text{ (exo)} \\ 1.59 \text{ (endo)} \end{cases}$	dt, $J_{7,8_{\text{exo}}} = J_{8_{\text{exo}},9} = 5, J_{8_{\text{exo}},8_{\text{endo}}} = 13$ d, $J_{8_{\text{exo}},8_{\text{endo}}} = 13$
9	83.3	4.31	t, $J_{8_{\text{exo}},9} = J_{9,10} = 5$
10	82.9	5.02	t, $J_{9,10} = J_{10,11} = 5$
11	63.1	3.88	dd, $J_{10,11} = 5, J_{6,11} = 10$
12	110.6 <sup>c</sup>	-	-
13	89.7 <sup>c</sup>	-	-
14	26.8	$\begin{cases} 2.08 \\ 2.33 \end{cases}$	sextet, $J_{14,14} = 14, J_{14,15} = 7$ sextet, $J_{14,14} = 14, J_{14,15} = 7$
15	10.0	1.06	t, $J_{14,15} = 7$

Table II.  $^{13}\text{C}^{\text{a}}$  and  $^1\text{H}^{\text{b}}$  NMR Data for Isomaneonene-B (2)

Carbon No.	$^{13}\text{C}$ $\delta$	$^1\text{H}$ $\delta$	Multiplicity, J (Hz)
1	83.5	2.72	d, $J_{1,3} = 2$
2	79.9 <sup>c,d</sup>	-	-
3	108.7	5.08	dd, $J_{1,3} = 2, J_{3,4} = 10.5$
4	146.9	6.34	dd, $J_{3,4} = 10.5, J_{4,5} = 9$
5	43.2	3.53	t, $J_{4,5} = J_{5,6} = 9$
6	50.0	2.75	sextet, $J_{5,6} = 9, J_{6,7} = 5, J_{6,11} = 10$
7	77.6	4.08	t, $J_{6,7} = J_{7,8_{\text{exo}}} = 5$
8	41.1	$\begin{cases} 1.15 \text{ (exo)} \\ 1.67 \text{ (endo)} \end{cases}$	dt, $J_{7,8_{\text{exo}}} = J_{8_{\text{exo}},9} = 5, J_{8_{\text{exo}},8_{\text{endo}}} = 13$ d, $J_{8_{\text{exo}},8_{\text{endo}}} = 13$
9	81.4	4.25	t, $J_{8_{\text{exo}},9} = J_{9,10} = 5$
10	84.6	5.03	t, $J_{9,10} = J_{10,11} = 5$
11	58.7	2.96	dd, $J_{10,11} = 5, J_{6,11} = 10$
12	109.5 <sup>c</sup>	-	-
13	85.2 <sup>c</sup>	-	-
14	38.0	$\begin{cases} 1.30 \\ 2.36 \end{cases}$	sextet, $J_{14,14} = 14, J_{14,15} = 7$ sextet, $J_{14,14} = 14, J_{14,15} = 7$
15	10.6	0.95	t, $J_{14,15} = 7$

Proton decoupled values (ppm downfield from TMS) in  $\text{CDCl}_3$  solvent. <sup>b</sup> 270 MHz values (ppm downfield from TMS) in  $\text{C}_6\text{D}_6$  solvent. <sup>c</sup> Quaternary carbon. <sup>d</sup> Reference 6.



### Acknowledgements

This work was supported by the National Institutes of Health (1 R01 CA16267). We are indebted to Drs. P. J. Scheuer and R. E. Moore, University of Hawaii, for the use of their laboratories during collecting trips, to Dr. M. R. Brennan, S.L., Dr. B. J. Burrenson, M. Serranon, and D. Dalietos for assistance in collecting the alga, to Mr. J. B. Keenaghan, Astra Pharmaceutical Products Inc., for the low resolution mass spectra, and to Dr. W. Anderson, Worcester Foundation for Experimental Biology, for the 100 MHz pmr spectra. The high resolution mass spectra were provided by the facility at MIT, Grant RR 9216 (Principal Investigator K. Biemann) and the FD mass spectra by the facility at Michigan State University, Grant RR 00480 (Principal Investigator C. C. Sweeley). The cmr spectra were obtained at the New England Area Research Facility, University of Connecticut Health Center, Grant RR 0639 (Principal Investigator J. Glasel). The 270 MHz pmr spectra were provided by the Yale facility, Grant 1 P07 PR 00798 (Principal Investigator M. Saunders).

### References and Notes

1. S. M. Waraszkiewicz, H. H. Sun, and K. L. Erickson, *Tetrahedron Letters*, 3021 (1976).
2. Complete experimental details will be published in a full paper.
3. D. Gagnaire and E. Payo-Subiza, *Bull. Soc. Chim. France*, 2627 (1963).
4. The numbering system shown is in keeping with that used for other members of this class of compounds.
5. H. H. Sun, S. M. Waraszkiewicz, and K. L. Erickson, *Tetrahedron Letters*, 585 (1976).
6. W. Fenical, *J. Am. Chem. Soc.*, 96, 5580 (1974).